REFERENCES

- Abad M, Ruiz C, Martínez D, Mosquera G, Sánchez JL. 1995. Seasonal variations of lipid classes and fatty acids in flat oyster, *Ostrea edulis*, from San Cibran (Galicia, Spain). Comparative Biochemistry and Physiology 110 C, 109–118.
- Alkanani T, Parrish CC, Thompson RJ, McKenzie CH. 2007. Role of fatty acids in cultured mussels, *Mytilus edulis*, grown in Notre Dame Bay, Newfoundland. Journal of Experimental Marine Biology and Ecology 348(1):33-45.
- Apeti DA, Kim Y, Lauenstein G, Tull J, and Warner R. 2014. Occurrence of Parasites and Diseases in Oysters and Mussels of the U.S. Coastal Waters. National Status and Trends, the Mussel Watch monitoring program. NOAA Technical Memorandum NOSS/NCCOS 182. Silver Spring, MD 51 pp.
- Bayne BL, Thompson RJ, Widdows J. 1973. Some effects of temperature and food on the rate of oxygen consumption by *Mytilus edulis* L. Effects of temperature on ectothermic organisms 181-193.
- Bayne BL, Bubel A, Gabbott PA, Livingstone D R, Lowe DM, Moore MN. (1982). Glycogen utilisation and gametogenesis in *Mytilus edulis* L. Marine Biology Letters 3:89-105.
- Buck BH, Thieltges DW, Walter U, Nehls G, Rosenthal H. 2005. Inshore–offshore comparison of parasite infestation in *Mytilus edulis*: Implications for open ocean aquaculture. Journal of Applied Ichthyology 21(2):107-13.
- Biggs CR, Lowerre-Barbieri SK, Erisman B. 2018. Reproductive resilience of an estuarine fish in the eye of a hurricane. Biol. Lett. 14(11):20180579
- Buda C, Dey I, Balogh N, Horvath LI, Maderspach K, Juhasz M, Yeo YK, Farkas T, 1994. Structural order of membranes and composition of phospholipids in fish brain cells during thermal acclimatization. *Proc. Natl. Acad. Sci. U.S.A.* 91: 8234-8238.
- Byron C, Link J, Costa-Pierce B, Bengtson D. (2011a). Calculating ecological carrying capacity of shellfish aquaculture using mass-balance modeling: Narragansett Bay, Rhode Island. Ecological Modelling 222(10):1743-55.
- Byron C, Link J, Costa-Pierce B, Bengtson D. (2011b). Modeling ecological carrying capacity of shellfish aquaculture in highly flushed temperate lagoons. Aquaculture 314(1):87-99.
- Camacho AP, González R, Fuentes J. 1991. Mussel culture in Galicia (N.W. Spain). Aquaculture 94(2):263-78.

- Canzonier W. 1972. *Cercaria tenuans*, larval trematode parasite of *Mytilus* and its significance in mussel culture. Aquaculture 1: 267-278.
- Carrington E (2002) Seasonal variation in the attachment strength of blue mussels: causes and consequences. Limnology and Oceanography, 47, 1723–1733.
- Chipperfield PNJ. (1953). Observations on the breeding and settlement of *Mytilus edulis* (L.) in British waters. Journal of the Marine Biological Association of the United Kingdom 32(2):449-76.
- Chu F.-L.E., Webb KL, Chen J. (1990). Seasonal changes of lipids and fatty acids in oyster tissues (*Crassostrea virginica*) and estuarine particulate matter. Comparative Biochemistry and Physiology 95A, 385–391.
- Clements JC, Hicks C, Tremblay R, Comeau LA. (2018) Elevated seawater temperature, not pCO2, negatively affects post-spawning adult mussels (*Mytilus edulis*) under food limitation. Conserv Physiol 6(1): cox078; doi:10.1093/conphys/cox078.
- Cooke SJ, Sack L, Franklin CE, Farrell AP, Beardall J, Wikelski M, Chown SL. 2013. What is conservation physiology? perspectives on an increasingly integrated and essential science. Conservation Physiology 1(1).
- Costa-Pierce BA. 2010. Sustainable ecological aquaculture systems: The need for a new social contract for aquaculture development. Marine Technology Society Journal 44(3):88-112.
- Coulthard HS. (1929). Growth of the sea mussel. Contr. Can. Biol. Fish 4, 123-136
- Cropp R and Gabric A. 2002. Ecosystem adaptation: do ecosystems maximize resilience? Ecology 83: 2019–2026.
- Dankers N and Zuidema DR. 1995. The role of the mussel (*Mytilus edulis* L.) and mussel culture in the Dutch Wadden Sea. Estuaries 18(1):71-80.
- Darriba S. 2017. Histopathological atlas: Marine bivalve molluscs. 1st ed. Intecmar. Xunta de Galicia (Consellería do Mar).
- Davenport J and Chen X. 1987. A comparison of methods for the assessment of condition in the mussel (*Mytilus edulis L*.). Journal of Molluscan Studies 53(3):293-7.
- Dey I, Buda C, Wiik T, Halver JE, Farkas T, 1993. Molecular and structural composition of phospholipid membranes in livers of marine and freshwater fish in relation to temperature. *Proc. Natl. Acad. Sci. U.S.A.* 90: 7498-7502.
- Dridi S, Romdhane MS, Elcafsi M. 2007. Seasonal variation in weight and biochemical composition of the pacific oyster, *Crassostrea gigas* in relation to the gametogenic

cycle and environmental conditions of the Bizert lagoon, Tunisia. Aquaculture 263(1):238-48.

- Duinker A, Håland L, Hovgaard P, Mortensen S. 2008. Gonad development and spawning in one and two year old mussels (*Mytilus edulis*) from Western Norway. Marine Biological Association of the United Kingdom. Journal of the Marine Biological Association of the United Kingdom 88(7):1465.
- Dumbauld BR, Ruesink JL, Rumrill SS. (2009). The ecological role of bivalve shellfish aquaculture in the estuarine environment: A review with application to oyster and clam culture in west coast (USA) estuaries. Aquaculture 290(3):196-223.
- Dynesius M, Hylander K, Nilsson C. 2009. High resilience of bryophyte assemblages in streamside compared to upland forests. Ecology 90(4):1042-54.
- Emmett B, Thompson K, Popham JD. (1987). The reproductive and energy storage cycles of two populations of *Mytilus edulis* (Linne) from British Columbia. Journal of Shellfish Research 6(1):29-36.
- Filgueira R, Guyondet T, Comeau LA, Tremblay R. (2016). Bivalve aquacultureenvironment interactions in the context of climate change. Global Change Biology 22(12):3901-13.
- Freites, L., Fernández-Reiriz, M.J., Labarta, U., 2003. Biochemical composition and energy content of the mussel *Mytilus galloprovincialis* of subtidal and rocky shore origin: Influence of environmental variables and source of mussel seed. Ciencias Marinas 29, 603–619.
- Gadner D, Riley JP. 1972. The component fatty acids of the lipids of some species of marine and freshwater molluscs. Journal of the Marine Biological Association of the United Kingdom 52, 827–832.
- Gabbot AP. 1983. Developmental and seasonal metabolic activities in marine molluscs. In: Hochachka, P.W. (Ed.), The Mollusca. : Environmental Biochemistry and Physiology, vol. 2. Academic Press, New York, pp. 165–217.
- Galaktionov K, Bustnes J, Bårdsen B, Wilson J, Nikolaev K, Sukhotin A, Skírnisson K, Saville D, Ivanov M, Regel K. 2014. Factors influencing the distribution of trematode larvae in blue mussels *Mytilus edulis* in the north Atlantic and arctic oceans. Mar Biol 162(1):193-206.
- Hall JM, Parrish CC, Thompson RJ, 2002. Eicosapentaenoic acid regulates scallop (*Placopecten magellanicus*) membrane fluidity in response to cold. Biol. Bull. 202: 201-203.
- Harris LG, Tyrrell MC (2001) Changing community states in the Gulf of Maine: synergism between invaders, overfishing and climate change. Biological Invasions, 3, 9–21.

- Incze LS, Lutz RA, Watling L. 1980. Relationships between effects of environmental temperature and seston on growth and mortality of *Mytilus edulis* in a temperate northern estuary. Marine Biology 57(3):147-56.
- Inglis, G.J., Hayden, B.J. & Ross, A.H. 2000. An overview of factors affecting the carrying capacity of coastal embayments for mussel culture. NIWA Client Report; CHC00/69 Project No. MFE00505. Christchurch, New Zealand, National Institute of Water and Atmospheric Research, Ltd. 31 pp.
- Jeffs AG, Holland RC, Hooker SH, Hayden BJ. 1999. Overview and bibliography of research on the Greenshell mussel, *Perna canaliculus*, from New Zealand waters. Journal of Shellfish Research;18(2):347–60.
- Jones SJ, Lima FP, Wethey DS. 2010. Rising environmental temperatures and biogeography: Poleward range contraction of the blue mussel, *Mytilus edulis* L., in the western Atlantic. Journal of Biogeography 37(12):2243-59.
- Kelly JR and Scheibling RE. 2012. Fatty acids as dietary tracers in benthic food webs. Marine Ecology Progress Series 446:1-22.
- Khan MA, Parrish CC, Shahidi F. 2006. Effects of environmental characteristics of aquaculture sites on the quality of cultivated Newfoundland blue mussels (*Mytilus edulis*). Journal of Agricultural and Food Chemistry 54(6):2236-41
- Laws EA and Archie JW. 1981. Appropriate use of regression analysis in marine biology. Marine Biology 65(1):13-6.
- LeBlanc N, Landry T, Davidson J, Tremblay R, McNiven M. (2010). The Effect of Elevated Water Temperature Stress on the Mussel *Mytilus Edulis* (L.), Survival and Genetic Characteristics. Canadian Technical Report of Fisheries and Aquatic Sciences, Ottawa, Canada, 2900: vii + 19p.
- Leonard GH, Bertness MD, Yund PO (1999) Crab predation, waterborne cues, and inducible defenses in the blue mussel, *Mytilus edulis*. Ecology, 80, 1–14.
- Lesser MP. 2016. Climate change stressors cause metabolic depression in the blue mussel, *Mytilus edulis*, from the Gulf of Maine. Limnology and Oceanography 61(5):1705-17.
- Li Y, and Smayda TJ. 1998. Temporal variability of chlorophyll in Narragansett Bay, 1973–1990. ICES Journal of Marine Science, 55: 661–667.
- Li Y, Qin JG, Abbott CA, Li XX, Benkendorff K. 2007. Synergistic impacts of heat shock and spawning on the physiology and immune health of *Crassostrea gigas*: an explanation for summer mortality in Pacific oysters. American Journal of Physiology - Regulatory Integrative and Comparative Physiology 293, R2353-R2362.

- Li Y, Qin JG, Li XX, Benkendorff K. 2009a. Monthly variation of condition index, energy reserves and antibacterial activity in Pacific oysters, *Crassostrea gigas*, in Stansbury (South Australia). Aquaculture 286, 64-71.
- Li Y, Qin JG, Li XX, Benkendorff K. 2009b. Spawning-dependent stress responses in Pacific oysters *Crassostrea gigas*: a simulated bacterial challenge in oysters. Aquaculture 293, 164-171.
- Li Y, Qin JG, Li XX, Benkendorff K. 2009c. Spawning-dependent stress response to food deprivation in Pacific oyster *Crassostrea gigas*. Aquaculture 286, 309-317.
- Logue J, de Vries A, Fodor E, Cossins A, 2000. Lipid compositional correlates of temperature-adaptive interspecific differences in membrane physical structure. *J. Exp. Biol.* 203: 2105-2115.
- Lowerre-Barbieri S, Crabtree L, Switzer T., Walters Burnsed S. and Guenther C. (2015) Assessing reproductive resilience: an example with South Atlantic red snapper *Lutjanus campechanus*. Marine Ecology Progress Series 526, 125–141.
- Maloy AP. (2001). Gametogenic cycles of marine mussels, *Mytilus edulis* and *Mytilus trossulus*, in Cobscook Bay, Maine. DigitalCommons@UMaine.
- Markowitz K, Williams J, Krause M. 2016. Development of quantitative PCR assay for detection of the trematode parasite *Proctoeces maculatus* in the blue mussel *Mytilus edulis*. Diseases of Aquatic Organisms 122(2):125-36.
- Marsden ID and Maclaren SR. 2010. Short-term study testing the resilience of an estuarine bivalve to macroalgal mats. Hydrobiologia 649(1):217-29.
- Martínez-Pita I, Sánchez-Lazo C, Ruíz-Jarabo I, Herrera M, Mancera JM. 2012. Biochemical composition, lipid classes, fatty acids and sexual hormones in the mussel *Mytilus galloprovincialis* from cultivated populations in south Spain. Aquaculture 358-359:274-83.
- Marty Y, Delaunay F, Moal J, Samain JF, 1992. Changes in the fatty acid composition of *Pecten maximus* (L.) during larval development. Journal of Experimental Marine Biology and Ecology 163, 221–234.
- McKenzie JD. 1986. The reproductive cycle of *Mytilus edulis* L. from Lough Foyle. Irish Naturalists Journal 22, 13–16.
- Murray LG, Newell CR, Seed R. 2007. Changes in the biodiversity of mussel assemblages induced by two methods of cultivation. Journal of Shellfish Research 26(1):153-62
- Narváez, M, Freites, L, Guevara, M, Mendoza, J, Guderley, H, Lodeiros, C.J, Salazar, G, 2008. Food availability and reproduction affects lipid and fatty acid composition of

the brown mussel, *Perna perna*, raised in suspension culture. Comparative Biochemistry and Physiology B 149, 293–302.

- Newell RIE, Hilbish TJ, Koehn RK, Newell CJ. 1982. Temporal variation in the reproductive cycle of *Mytilus edulis* L. (Bivalvia, Mytilidae) from localities on the east coast of the United States. Biological Bulletin 162(3):299-310.
- NOAA National Marine Fisheries Service (2016) Commercial fisheries statistics: annual commercial landing statistics. Available at: http://www.st.nmfs.noaa.-gov/st1/commercial/landings/annual_landings.html
- Ojea J, Pazos AJ, Martínez D, Novoa S, Sánchez JL, Abad M, 2004. Seasonal variation in weight and biochemical composition of the tissues of *Ruditapes decussatus* in relation to the gametogenic cycle. Aquaculture 238, 451–468.
- Ortiz-Zarragoitia M and Cajaraville MP. 2010. Intersex and oocyte atresia in a mussel population from the biosphere's Reserve of Urdaibai (Bay of Biscay). Ecotoxicology and Environmental Safety 73(5):693-701.
- Parrish CC. 1987. Separation of aquatic lipid classes by chromarod thin layer chromatography with measurements by iatroscan flame ionization detection. Can. J. Fish. Aquat. Sci. 44: 722–731
- Parrish CC. 1999. Determination of total lipid, lipid classes and fatty acids in aquatic samples, p. 4–20. In M. T. Arts and B. C. Wainman [eds.], Lipids in freshwater ecosystems. Springer.
- Parrish CC, 2013. Lipids in marine ecosystems. ISRN Oceanography 2013:1-16.
- Pazos, AJ, Román, G, Acosta, CP, Sánchez, JL, Abad, M, 1997. Lipid classes and fatty acid composition in the female gonad of *Pecten maximus* in relation to reproductive cycle and environmental variables. Comparative Biochemistry and Physiology B 117, 393–402.
- Pazos, AJ, Sánchez, JL, Román, G, Pérez-Perellé, ML, Abad, M, 2003. Seasonal changes in lipid classes and fatty acid composition in the digestive gland of *Pecten maximus*. Comparative Biochemistry and Physiology 134B, 367–380.
- Pernet F, Gauthier-Clerc S, Mayrand E. 2007. Change in lipid composition in eastern oyster (*Crassostrea virginica* Gmelin) exposed to constant or fluctuating temperature regimes. *Comp. Biochem. Physiol.* 147: 557-565.
- Pershing AJ, Alexander MA, Hernandez CM, Kerr LA, Le Bris A, Mills KE, Nye JA, Record NR, Scannell HA, Scott JD, et al. 2015. Slow adaptation in the face of rapid warming leads to collapse of the Gulf of Maine cod fishery. Science 350(6262):809-12.

- Petes, LE, Menge BA, Harris, AL 2008. Intertidal mussels exhibit energetic trade-offs between reproduction and stress resistance. Ecological Monographs 78, 387-402.
- Pollero RJ, Re MR, Brenner RR. 1979. Seasonal changes of the lipids of the mollusc *Chlamys tehuelca*. Comparative Biochemistry and Physiology A 64, 257–263.
- Read KRH and Cummings KB. 1967. Thermal tolerance of the bivalve molluscs *Modiolus modiolus* (L.), *Mytilus edulis* L., and *Brachidontes demissus*. Comp. Biochem. Physiol. 22, 149-155.
- Ricklefs RE and Wikelski M. (2002) The physiology/life-history nexus. Trends Ecol. Evol. 17, 462–468
- Rowley AF, Cross ME, Culloty SC, Lynch SA, Mackenzie CL, Morgan E, O'Riordan RM, Robins PE, Smith AL, Thrupp TJ, et al. (2014). The potential impact of climate change on the infectious diseases of commercially important shellfish populations in the Irish sea—a review. ICES Journal of Marine Science 71(4):741-59.
- Schulte EH. (1975). Influence of algal concentration and temperature on the filtration rate of *Mytilus edulis*. Marine Biology, 30, 331–341.
- Seed R. (1969). The ecology of *Mytilus edulis* L. (lamellibranchiata) on exposed rocky shores. I. breeding and settlement. Oecologia 3(3/4):277-316.
- Sokolova IM, Frederich M, Bagwe R, Lannig G, Sukhotin AA. 2012. Energy homeostasis as an integrative tool for assessing limits of environmental stress tolerance in aquatic invertebrates. Marine Environmental Research 79:1-15.
- Song L, Li X, Clarke S, Wang T, Bott K, 2007. The application of neutral red retention assay to evaluate the differences in stress responses to sexual maturation and spawning between different sizes of Pacific oyster, *Crassostrea gigas* (Thunberg). Journal of Shellfish Research 26, 493-499.
- Sorte CJB, Etter RJ, Spackman R, Boyle EE, Hannigan RE. 2013. Elemental fingerprinting of mussel shells to predict population sources and redistribution potential in the Gulf of Maine. PloS One 8(11).
- Sorte CJB, Davidson VE, Franklin MC, Benes KM, Doellman MM, Etter RJ, Hannigan RE, Lubchenco J, Menge BA. (2016). Long- term declines in an intertidal foundation species parallel shifts in community composition. Global Change Biology 23(1):341-52.
- Soudant, P., Marty, Y., Moal, J., Robert, R., Quere, C., Lecoz, J.R., Samain, J.F., 1996a. Effect of food fatty acid and sterol quality on *Pecten maximus* gonad composition and reproduction process. Aquaculture 143, 361–378.

- Soudant, P., Marty, Y., Moal, J., Robert, R., Quere, C., Lecoz, J.R., Samain, J.F., 1996b. Impact of the quality of dietary fatty acids on metabolism and the composition of polar lipid classes in female gonads of *Pecten maximus* (L.). Journal of Experimental Marine Biology and Ecology 205, 149–163.
- Soudant, P., Van Ryckeghem, K., Marty, Y., Moal, J., Samain, J.F., Sorgeloos, P., 1999. Comparison of the lipid class and fatty acid compositions between a reproductive cycle in nature and a standard hatchery conditioning of the pacific oyster *Crassostrea* gigas. Comparative Biochemistry and Physiology B 123, 209–222.
- Stunkard, H. W., and Uzmann J. R. (1959). The life-cycle of the digenetic trematode, *Proctoeces maculatus* (Looss, 1901) Odhner, 1911 [syn. *P. subtenuis* (Linton, 1907) Hanson, 1950], and description of *Cercaria adranocerca* n. sp. Biological Bulletin 116(1):184-93.
- Suárez M. P., Alvarez C., Molist P., San Juan F. (2005). Particular aspects of gonadal cycle and seasonal distribution of gametogenic stages of *Mytilus galloprovincialis* cultured in the estuary of Vigo. Journal of Shellfish Research 24(2):531-40.
- Sunila I, Williams L, Russo S, Getchis T. (2004). Reproduction and pathology of blue mussels, *Mytilus edulis* in an experimental longline in long island sound, Connecticut. Journal of Shellfish Research 23(3):731.
- Swift, M.L., 1977. Phosphono-lipid content of the oyster, *Crassostrea virginica*, in three physiological conditions. Lipids 15, 129–132.
- Tate RD, Benkendorff K, AbLah R, Kelaher BP (2017) Ocean acidification and warming impacts the nutritional properties of the predatory whelk *Dicathais orbita*.J Exp Mar Biol Ecol 493: 7–13.
- Thieltges D, Hussel B, Hermann J, Jensen K, Krakau M, Taraschewski H, Reise K. 2008. Parasites in the northern Wadden sea: A conservative ecosystem component over 4 decades. Helgol Mar Res 62(1):37-47.
- Thrush SF, Haliday J, Hewitt JE & Lohrer AM. 2008. The effects of habitat loss, fragmentation, and community homogenisation on resilience in estuaries. Ecological Applications 18: 12–21.
- Tiku PE, Gracey AY, Macartney AI, Beynon RJ, Cossins AR. 1996. Cold-induced expression of delta0-desaturase in carp by transcriptional and posttranslational mechanisms. Science 271: 815-818.
- Valles-Regino R, Tate R, Kelaher B, Savins D, Dowell A, Benkendorff K. (2015) Ocean warming and CO2-induced acidification impact the lipid content of a marine predatory gastropod. Mar Drugs 13: 6019–6037.

- Wardle WJ. (1980). On the life cycle stages of *Proctoeces maculatus* (Digenea: Fellodistomidae) in mussels and fishes from Galveston bay, Texas. Bulletin of Marine Science 30(3):737-43.
- Widdows J. 1973a. Effect of temperature and food on the heart beat, ventilation rate and oxygen uptake of *Mytilus edulis*. Marine Biology 20(4):269-76.
- Widdows J. 1973b. The effects of temperature on the metabolism and activity of *Mytilus edulis*. Netherlands Journal of Sea Research 7:387-98.
- Wikelski M, Cooke SJ (2006) Conservation physiology. Trends Ecol Evol 21: 38-46.
- Winstead JT. 1995. Digestive tubule atrophy in Eastern oysters, *Crassostrea virginica* (Gmelin, 1791), exposed to salinity and starvation stress. J. Shellfish Res., 14:105–111.
- Zhukova NV, Kharlamenko VI, Svestashev VI, Rodionov IA, 1992. Fatty acids as markers of bacterial symbionts of marine bivalve molluscs. J. Exp. Mar. Biol. Ecol. 162, 253–263.

FIGURES



Figure 1. Map of Casco Bay. At left, map of the sampling area, Casco Bay, Maine. The exact locations of the farms have been left out for the sake of keeping our partner farm anonymous. Inlay shows the Northeast Coast of the United States with Casco Bay boxed in black.



Figure 2. Study Site Environmental Conditions. Average monthly water temperature (red) and chlorophyll concentration (green) for the duration of this study, in Casco Bay. Standard Deviation bars shown.

















Figure 3. Staging Scale. Created for this study modified from Chipperfield 1953 and Duinker et al. 2008. 'FM0' denotes Stage 0. 'F1.-F5.' represent female stages 1-5. 'M1.-M4.' represent male stages 1-5.



Figure 4. Pathology Micrographs. Examples of the five pathologies of focus for this study. (a.) Black arrow pointing to gill ciliates in the gills of a mussel. (b.) Digestive gland atrophy (DGA), black arrows pointing out tubules with thinned walls. (c.) Oocyte atresia. (d.) Hemocyte filled mantle follicles. (e. & f.) Trematode sporocysts containing cercaria, adjacent to digestive tubules (e.) and in the mantle (f.).



Figure 5. Fatty Acid Principal Coordinates Analysis. Principal coordinates analysis describing the degree of similarity between samples based on their fatty acid composition (in terms of percentage per sample, not wet weight). Symbols represent sampling dates, each sampling date has three replicate mussel samples. Samples within the green (75) circle are at least 75% similar in fatty acid composition. All samples are at least this similar as they are all the same species, sampled in close proximity to each other. Samples within the dotted blue (80) circles are at least 80% similar in fatty acid composition. This is split into two groups that can roughly be classified as Summer & Fall samples (left) and Winter & Spring samples (right). Samples within the dotted light blue (90) circles are at least 90% similar in their fatty acid composition. These circles represent samples taken on the same day or similar days. Indicating that these mussels were all of similar condition and feeding on similar items.



Figure 6. Condition Index and Energy Investment. Average percentage of the mantle occupied by gonad tissue (purple line), vesicular connective tissue (VCT, glycogen) (gray line) and adipogranular tissue (ADG, protein, lipid, glycogen) (red line) for each month of this study with standard deviation shown. Average Condition Index for each month represented by gray bars. Sexes combined for each line/bar. (SD bars: gonad in purple, VCT in gray, ADG in red, CI in gold)



Figure 7. Female Staging Results. Female mussels, percentage of each maturity stage, per month. Number of mussels analyzed each month shown above bar.



Figure 8. Male Staging Results. Male mussels, percentage of each maturity stage, per month. Number of mussels analyzed each month shown above bar.



Figure 9. Lipids and Energy Investment. Average wet weights $(\mu g/g)$ for select lipids. The most abundant lipid classes were chosen for this figure. The gray bars (total lipids) represents an average wet weight all of the lipids detected for each month. SD bars shown. Overlaid are the curves of average percentage of the mantle occupied by gonad tissue (purple line) and adipogranular tissue (ADG, protein, lipid, glycogen) (red line) for each month of this study.



Figure 10. Fatty Acids and Chlorophyll Concentration. Average wet weights $(\mu g/g)$ for select fatty acid biomarkers. SD bars shown. Biomarkers were chosen based on the main components of a blue mussel's diet; dinoflagellates, bacteria, diatoms and detritus. Average monthly chlorophyll concentration represented by black line.



Figure 11. Fatty Acids and Energy Investment. Average wet weights $(\mu g/g)$ for select fatty acid biomarkers. Biomarkers were chosen based on the main components of a blue mussel's diet; dinoflagellates, bacteria, diatoms and detritus. Overlaid are the curves of average percentage of the mantle occupied by gonad tissue (purple line) and adipogranular tissue (ADG, protein, lipid, glycogen) (red line) for each month of this study.



Figure 12. Condition Index and Storage: Gonad. Average Condition Index for each month (gray bars) and ratio of Storage tissue (VCT and ADG) to Gonad tissue (yellow bars).



Figure 13. Female Staging Results vs. Temperature. Female staging results plotted versus average water temperature from the week preceding the sampling event. All sampling events for the duration of this study are shown (n=43). The reproductive cycle, as it relates to water temperature can be broken down into three distinct periods. From $-1.1^{\circ}C - 9.1^{\circ}C$ the mussels are in a developing period characterized by high levels of Stage 1 and 2 follicles, with Stage 3 follicles increasing in percentage as temperature increases. From $9.2^{\circ}C - 12.3^{\circ}C$ the mussels are mature, nearly 90% of the follicles from these sampling dates were filled with mature and ready to be spawned oocytes. At this point there was little presence of Stage 4 follicles, indicating that mussels had not begun large spawning efforts. The third period, spawning and redeveloping, occurs between $13^{\circ}C - 19.2^{\circ}C$. This is when the highest percentage of post spawning stages are observed. Many lower stages are observed during these temperatures as well, indicating mussels that may be redeveloping after spawning in preparation for another spawning event.



Figure 14. Energy Investment vs. Temperature. Average percentage of the mantle occupied by gonad tissue (purple dots), vesicular connective tissue (VCT, glycogen) (gray dots) and adipogranular tissue (ADG, protein, lipid, glycogen) (red dots) versus the average water temperature of the week preceding the sampling event.

TABLES

Food Source	FA biomarkers used
Diatoms	16:1ω7, 20:5ω3 (EPA)
Dinoflagellates	22:6ω3 (DHA)
Detritus (macroalgae, salt marsh plants, seagrass)	18:1009, 18:2006, 18:3003, 18:4003, 20:4006, 22:0
Bacteria	i15:0, ai15:0, i17:0, ai17:0, 18:1ω7

Table 1. Fatty Acid Biomarkers. Fatty acid biomarkers used in this study, assigned to the main components of a mussel's diet. Based on Kelly & Scheibling 2012 and Parrish 2013.

	r	rsquare
gonad:ADG	-0.7	0.5
gonad:VCT	-0.7	0.4
gonad:temp	-0.2	0.0
gonad:chla	0.0	0.0
ADG:VCT	0.5	0.2
ADG:temp	-0.1	0.0
ADG:chla	0.1	0.0
VCT:temp	0.2	0.0
VCT:chla	-0.1	0.0

Table 2. Linear Regressions. Correlation coefficients and coefficients of determination based on Model II linear regressions run between components of energy investment and environmental conditions. n for each regression was 375.

		201	7	2018				
month	avg. temp	SD (+/-)	avg. chl	SD (+/-)	avg. temp	SD (+/-)	avg. chl	SD (+/-)
Jan	\	\	\	\	0.0	0.8	12.6	4.2
Feb	1.7	1.3	3.9	2.5	2.0	1.1	7.2	3.3
Mar	2.0	0.8	10.7	5.6	3.5	0.7	24.9	22.8
Apr	6.2	1.9	5.7	5.4	5.6	1.2	15.5	14.9
May	9.5	1.5	2.7	1.4	10.9	1.2	2.1	1.5
Jun	13.3	1.7	2.5	2.1	13.6	1.1	3.2	1.2
Jul	16.2	1.4	3.3	2.9	17.1	1.5	4.2	2.2
Aug	16.9	1.2	8.0	4.5	18.7	1.3	5.7	3.7
Sept	15.5	0.9	5.6	5.9	17.2	1.2	4.6	2.3
Oct	14.1	0.8	7.4	4.3	15.4	2.1	4.8	2.2
Nov	8.9	2.1	8.5	5.4	\	\	\	\
Dec	3.9	2.3	9.4	4.2	\	\	\	\

Table 3. Environmental Data Summary. Average environmental conditions for each month of this study and their corresponding standard deviations.

		Digestive Gland				Hemocyte Filled						
		Atrophy		Oocyte Atresia		Mantle P	Mantle Follicles		Gill Ciliates		Trematodes	
year	month	Prevalence	Intensity	Prevalence	Intensity	Prevalence	Intensity	Prevalence	Intensity	Prevalence	Intensity	
2017	Feb	0.0	0.0	0.8	1.0	0.2	0.5	0.3	1.5	0.0	١	
2017	Mar	0.0	0.0	1.0	2.0	0.4	0.5	0.0	0.0	0.0	\	
2017	Apr	0.2	0.8	1.0	2.1	0.1	1.0	0.1	1.0	0.0	\	
2017	May	0.3	0.8	1.0	1.8	0.2	1.0	0.0	0.0	0.0	١	
2017	Jun	0.4	0.8	1.0	2.1	0.4	4.5	0.1	1.0	0.0	١	
2017	Jul	0.6	1.2	0.9	2.4	0.5	4.0	0.2	1.6	0.0	١	
2017	Aug	0.6	1.0	0.9	2.4	0.5	5.3	0.1	1.5	0.1	١	
2017	Sept	0.5	1.5	0.9	2.8	0.3	5.8	0.2	1.3	0.0	١	
2017	Oct	0.5	1.0	0.6	2.3	0.4	3.3	0.2	0.9	0.0	١	
2017	Nov	0.3	1.8	0.2	0.5	0.2	2.0	0.1	1.0	0.0	١	
2017	Dec	0.1	1.0	0.8	1.4	0.4	3.2	0.1	1.0	0.0	١	
2018	Jan	0.1	0.3	0.8	2.1	0.6	6.4	0.1	0.7	0.1	١	
2018	Feb	0.1	0.2	1.0	2.3	0.6	0.9	0.2	0.7	0.0	١	
2018	Mar	0.4	1.0	0.9	1.8	0.4	1.5	0.2	1.0	0.0	١	
2018	Apr	0.8	1.3	1.0	1.6	0.2	1.0	0.1	1.0	0.0	١	
2018	May	1.0	1.8	1.0	3.3	0.2	1.7	0.2	0.8	0.0	١	
2018	Jun	1.0	1.6	1.0	2.6	0.3	1.5	0.2	1.8	0.0	١	
2018	Jul	1.0	2.0	0.9	2.7	0.5	4.5	0.5	2.2	0.0	١	
2018	Aug	1.0	2.1	0.8	2.5	0.1	2.1	0.1	0.0	0.0	١	
2018	Sept	0.9	2.2	0.9	1.9	0.1	2.2	0.2	1.5	0.0	١	
2018	Oct	0.9	1.8	0.9	0.9	0.0	1.8	0.1	1.5	0.0	١	

Table 4. Pathological Survey Summary. Results of the pathological survey. Presence and intensity of each factor for each month of this study. Because only five mussels were found to contain trematodes, intensities were not calculated. Intensities for digestive gland atrophy and oocyte atresia are on a scale of 0-4; 0 being no detection of condition, 4 being the condition is detected in over 75% of the target tissue.