

Improvement of Grafted Watermelon Transplant Survival as a Result of Size and Starch Increases Over Time Caused by Rootstock Fatty Alcohol Treatment: Part II

Shawna L. Daley^{1,3}, William Patrick Wechter², and Richard L. Hassell¹

ADDITIONAL INDEX WORDS. regrowth control, rootstock treatment, vegetable grafting, watermelon grafting

SUMMARY. Fatty alcohol treatments can be used to eliminate the meristem of cucurbit (Cucurbitaceae) rootstocks, which prevents regrowth when grafting, but the effects of the treatment on the rootstock have not been documented. Two rootstock types, ‘Emphasis’ bottle gourd (*Lagenaria siceraria*) and ‘Carnivor’ interspecific hybrid squash (*Cucurbita maxima* × *C. moschata*) commonly used in watermelon (*Citrullus lanatus*) grafting significantly increased in cotyledon and hypocotyl size over 21 days after treatment (DAT) with a 6.25% fatty alcohol emulsion. There was a significant increase in total soluble sugar (glucose, sucrose, and fructose) content for each rootstock hypocotyl and cotyledon. Starch concentrations of hypocotyls and cotyledons also increased significantly in both rootstocks. This increase in stored energy could greatly increase the success rate of the grafting process. Increased rootstock energy reserves could overcome the need for keeping the rootstock cotyledon intact when grafting.

Grafting onto disease-resistant rootstocks is an important technology for overcoming soil-borne disease (Buller et al., 2013; Guan et al., 2012; Louws et al., 2010) and is used for watermelon production primarily in Asia, Europe, and the Middle East (Cohen et al., 2007; Davis et al., 2008). However, the high cost of grafted transplants has prevented the use of grafted transplants in U.S. production. As methyl bromide, an effective and inexpensive soil fumigant, has been phased out (U.S. Department of Agriculture, 2012) and disease-free land has become scarce, there has begun to be a need for grafted watermelon transplants in the United States. With current disease conditions in U.S. soils, it is estimated that at least 6.69 million grafted watermelon transplants per year would be required to combat soil-borne diseases previously controlled with fumigants (D. Liere, personal communication).

Much of the cost of grafted transplants is associated with the labor involved in their production and maintenance (Davis et al., 2008). The majority of these labor costs are a result of rootstock regrowth (Choi et al., 2002; Memmott and Hassell, 2010), which competes with the watermelon scion for nutrients and sunlight. In countries currently using grafted transplants commercially, the meristem is manually removed at the grafting stage (Hassell et al., 2008). Meristematic regrowth is manually removed during graft healing, at the transplanting stages, and thereafter as needed (Lee and Oda, 2003). Rootstock regrowth removal is an expensive, labor-intensive process and has been prohibitive to the adoption of grafted transplants in U.S. watermelon production.

Rootstock treatment with fatty alcohol can control meristematic

regrowth by burning the meristem tissue and allow for successful grafting without the risk of regrowth. Commercial watermelon grafting methods require at least one rootstock cotyledon to remain intact to ensure graft success, and grafting must be done soon after the cotyledons unfold to limit regrowth (Hassell et al., 2008; Lee and Oda, 2003). Rootstock cotyledons provide energy to maintain the plant and heal the graft. Cucurbit seedlings are dependent upon seed reserves only during preemergent growth (Bisognin et al., 2005) and depend completely on the leaf-like, highly photosynthetic cotyledons for further development (Penny et al., 1976). At least one rootstock cotyledon is needed for early growth and establishment (Bisognin et al., 2005).

Fatty alcohol was originally used to control growth of axillary meristems on topped tobacco (*Nicotiana tabacum*). Because the fatty alcohols target only the actively dividing cells in meristems, the tobacco leaves remain intact, and energy that would have been sent to new growth is now stored in the leaves (Steffens et al., 1967). An application of fatty alcohol to the apical meristem of cucurbit rootstock seedlings before grafting has been shown to be an effective means of eliminating meristem tissue and controlling regrowth (Daley and Hassell, 2014). We observed that over a 21-d period following fatty alcohol treatment, both bottle gourd and interspecific hybrid squash rootstock seedling cotyledons seemed to expand, becoming long and rigid. The hypocotyl also increased in length and diameter over the observed time, yet the seedlings remained viable for grafting. We hypothesize that the increase in seedling size was due to an accumulation of carbohydrates within the rootstock. Thus, this experiment was designed to determine changes in rootstock seedling development over

Technical Contribution No. 6262 of the Clemson University Experiment Station.

¹Department of Plant and Environmental Sciences, Clemson University Coastal Research and Education Center, 2700 Savannah Highway, Charleston, SC 29414

²USDA-Agriculture Research Service, 2700 Savannah Highway, Charleston, SC 29414

³Corresponding author. E-mail: sdaley@clemson.edu.

Units

To convert U.S. to SI, multiply by	U.S. unit	SI unit	To convert SI to U.S., multiply by
29,574	fl oz	μL	3.3814×10^{-5}
2.54	inch(es)	cm	0.3937
25.4	inch(es)	mm	0.0394
0.2276	inch ² /oz	cm ² ·g ⁻¹	4.3942
0.0254	mil	mm	39.3701
28.3495	oz	g	0.0353
1	ppm	μg·g ⁻¹	1
$(F - 32) \div 1.8$	°F	°C	$(C \times 1.8) + 32$

21 DAT with fatty alcohol, as well as the effect on carbohydrate concentrations within the hypocotyls and cotyledons of the rootstock seedlings.

Materials and methods

PLANT MATERIAL AND GROWING CONDITIONS. Two rootstock types were chosen: ‘Emphasis’ bottle gourd (Syngenta Seeds, Boise, ID) and ‘Carnivor’ interspecific hybrid squash (Syngenta Seeds). Seeds were sown in 72-cell plug trays with a 1-inch diameter (TLC Polyform, Minneapolis, MN) using a fertilizer-free, soil-less mix (76% sphagnum peat, 25% perlite; Sun Gro Horticulture, Agawam, MA). No additional fertilizer was applied throughout the experiment. Seeds were planted following standard greenhouse production practices (Rutledge, 2009). Plants were grown in a greenhouse covered with a double-layer of 6-mil polyethylene (K50 Clear; Klerks Hyplast, Chester SC). The minimum temperature was set at 60 °F, and exhaust fans were set to power on at 70 °F.

MERISTEM TREATMENT. When cotyledons had unfolded, but not fully expanded (\approx 8–10 d after seeding), plants were acclimated in a climate-controlled room (75 °F, 70% to 75% relative humidity) for 12 h before treatment. Seedlings were then treated with 20 μ L of a 6.25% fatty alcohol emulsion (Fair 85[®], Fair Products, Cary, NC), which had previously been determined to be the optimal treatment volume and concentration to avoid regrowth and seedling mortality (Daley and Hassell, 2014). The fatty alcohol emulsion was applied with a single-channel micropipette (VWR International Radnor, PA) to the apical meristem area of each seedling. Water-only control seedlings were treated in the same manner with 20 μ L distilled water. Trays remained in the treatment room for 5 h to allow the fatty alcohol or water treatments, respectively, to penetrate leaf tissue. After treatment penetration, seedlings were returned to the greenhouse for the duration of the experiment. While in the greenhouse, rootstocks were checked daily and any regrowth removed.

EXPERIMENTAL DESIGN. The experiment was arranged in a completely randomized design, with 72 plants randomly selected from five trays for each rootstock–time treatment

combination. The experiment was repeated twice. Carbohydrate analysis was also a completely randomized design with 12 samples per treatment. Each sample was a composite of either 5 hypocotyls or 10 cotyledons.

VISIBLE PLANT ANALYSIS. On 1, 7, 14, and 21 DAT, 72 rootstock seedlings from the rootstock–fatty alcohol treatment were destructively harvested and measured. The water-only control seedlings were only harvested 1 d following fatty alcohol application. A razor blade was used to separate the hypocotyl from the roots at the soil surface. The cotyledons were removed at the point of attachment to the hypocotyl using the same blade. Individual hypocotyl and cotyledon fresh weights (Sartorius A 120 S; Data Weighing Systems, Elk Grove, IL), and length and width of all 72 seedlings were measured with a digital caliper (CD-6 B; Mitutoyo America Corp., Aurora, IL). Cotyledon leaf area was measured using a leaf area meter (LI 3100; LICOR, Lincoln, NE). After measurements were recorded, hypocotyls and cotyledons were dried separately for 48–72 h at 70 °F (320–6 1000 Series; Napco Industrial Partner Richardson, TX), and dry weights of individual hypocotyls and cotyledons were recorded. Tissues of each organ were combined and then randomly divided into 12 samples, either 5 hypocotyls or 10 cotyledons, for carbohydrate analysis. Each of the 12 samples was ground using a mortar and pestle and stored at –20 °C until analyzed.

CARBOHYDRATE EXTRACTION. Glucose, sucrose, fructose, and starch were enzymatically extracted from 70 mg dried, ground tissue following the protocol of Zhao et al. (2010). Microplate enzymatic assays were carried out to detect each sugar (Zhao et al., 2010). Sugar concentrations were calculated using absorbance, volume sampled, and sample weight. Mean carbohydrate concentrations were then multiplied by the average dry weight per organ for each time after treatment to determine the amount of carbohydrate available in an average hypocotyl or cotyledon. Glucose, sucrose, and fructose results were combined and are reported as total soluble sugars, and starch results are reported alone.

STATISTICAL ANALYSIS. Seedling growth measurements and carbohydrate

data were analyzed using a linear mixed model in the fit model platform of JMP Pro 10 (SAS Institute, Cary, NC). The model was a complete factorial model of interactions between the fixed effect of DAT and the random effect of planting date. As rootstocks are of different species, each rootstock–organ combination was analyzed separately. Mean comparisons were performed using Fisher’s protected least significant difference test, with $P \leq 0.05$.

Results

VISIBLE PLANT ANALYSIS. For both rootstock cultivars, analysis of variance revealed significant effects of DAT for rootstock hypocotyl fresh and dry weights and width (Table 1). In both rootstock types, hypocotyl development measurements (fresh weight, dry weight, length, and width) at 1 DAT were statistically similar to the measurements of the water-treated control, indicating that the fatty alcohol treatment had no detrimental effect on rootstock development (Table 1). Mean hypocotyl fresh weights of both ‘Emphasis’ and ‘Carnivor’ rootstocks increased significantly (2.6- and 2.0-fold, respectively) over 21 DAT (Table 1). ‘Emphasis’ hypocotyl dry weights increased 4.4-fold, and ‘Carnivor’ hypocotyl dry weights increased 5.2-fold, over 21 DAT. ‘Emphasis’ hypocotyl length did not significantly change over the course of the experiment, but ‘Carnivor’ hypocotyl lengths decreased significantly between 1 and 7 DAT. ‘Emphasis’ hypocotyl width increased 1.6-fold between 7 and 21 DAT, and ‘Carnivor’ hypocotyl width increased 1.5-fold.

Similar to rootstock hypocotyl development, for both rootstocks, there was a significant effect of DAT in cotyledon fresh and dry weights, as well as ‘Carnivor’ cotyledon width, indicating that the rootstock seedlings increased in size after meristem removal (Table 2). Between 7 and 21 DAT, fresh weights of ‘Emphasis’ and ‘Carnivor’ rootstocks increased by 1.5- and 1.4-fold, respectively (Table 2). ‘Emphasis’ rootstock cotyledon dry weight increased by 2.6-fold 7 DAT compared with 1 DAT, and cotyledon dry weight did not significantly increase on subsequent days. However, ‘Carnivor’ rootstock cotyledon dry

weight increased 3-fold between 1 and 21 DAT. 'Emphasis' cotyledon length increased by 7.36 mm between 7 and 21 DAT, and 'Carnivor' cotyledon length increased by 9.24 mm between 1 and 21 DAT. 'Emphasis' cotyledon width and thickness did not change significantly over the course of the experiment. 'Carnivor' cotyledon width increased by 5.5 mm between 1 and 7 DAT, and cotyledon thickness estimates increased by 0.01 cm²·g⁻¹ between 7 and 14 DAT.

CARBOHYDRATE ANALYSIS. There was a significant effect of days on both total soluble sugars and starch content in hypocotyls and cotyledons of 'Carnivor' and 'Emphasis' rootstock cultivars, with the exception of starch content in 'Emphasis' hypocotyls, indicating that rootstocks increase in total soluble sugar and starch content over time after fatty alcohol treatment (Table 3).

With both rootstock cultivars, there were no significant differences in total soluble sugars or starch

content between the rootstocks treated with water and with fatty alcohol 1 DAT, indicating that fatty alcohol treatment does not affect baseline amounts of total soluble sugars or starch (Table 3). Total soluble sugar content of 'Emphasis' and 'Carnivor' rootstock hypocotyl increased 2.5- and 6.7-fold, respectively, over the course of the experiment. Starch content of 'Emphasis' hypocotyls increased by 31.5-fold by 21 DAT, although the increase was not significant. Starch content of 'Carnivor' hypocotyls significantly increased 193.6-fold at 21 DAT. 'Emphasis' cotyledons increased in total soluble sugar content by 8.2-fold between 1 and 14 DAT, and did not increase significantly between 14 and 21 DAT. Similarly, 'Carnivor' cotyledons increased in total soluble sugar content by 3.5-fold between 1 and 7 DAT, and increased by 1.4-fold between 7 and 21 DAT. Starch content in 'Emphasis' cotyledons increased 228.7-fold between 1 and 7 DAT, and increased 2.2-fold between 7 and 21 DAT. 'Carnivor' cotyledons increased in starch content 29.2-fold between 1 and 7 DAT, and by 1.9 fold between 14 and 21 DAT.

Discussion

We observed no significant differences in rootstock development 1 DAT with water or fatty alcohol. Similarly, there were no significant differences in total soluble sugars or

Table 1. Effect of time after water and fatty alcohol treatment on bottle gourd 'Emphasis' and interspecific hybrid squash 'Carnivor' rootstock hypocotyl development.

Rootstock	DAT ^z	Hypocotyl ^y			
		Fresh wt (g) ^x	Dry wt (g)	Length (mm) ^x	Width (mm)
'Emphasis'	1 (water)	0.23 c ^w	0.013 d	36.87	2.63 c
	1	0.23 c	0.013 d	36.41	2.53 c
	7	0.30 c	0.023 c	34.97	3.06 c
	14	0.48 b	0.040 b	35.82	4.18 b
	21	0.62 a	0.056 a	37.48	4.99 a
<i>P</i> value		***	***	NS	**
'Carnivor'	1 (water)	0.37 c	0.025 c	54.20 a	2.80 c
	1	0.35 c	0.023 c	51.65 a	2.84 c
	7	0.44 bc	0.055 c	42.95 b	3.55 b
	14	0.59 ab	0.090 b	41.55 b	4.22 a
	21	0.75 a	0.134 a	41.39 b	4.48 a
<i>P</i> value		*	**	**	**

^zDays after treatment with fatty alcohol or water. All rootstocks except the water control treatments were treated with a 6.25% fatty alcohol emulsion.

^ySingle hypocotyl measurements.

^x1 g = 0.0353 oz, 1 mm = 0.0394 inch.

^wMeans within cultivar and column having the same or no letters are not significantly different by Fishers' protected least significant difference test; NS, *, **, *** (not significant or significant at $P \leq 0.05$, 0.01, and 0.001).

Table 2. Effect of time after water and fatty alcohol treatment on 'Emphasis' bottle gourd and 'Carnivor' interspecific hybrid squash rootstock cotyledon development.

Rootstock	DAT ^z	Cotyledon ^y				
		Fresh wt (g) ^x	Dry wt (g)	Length (mm) ^x	Width (mm)	Thickness (cm ² ·g ⁻¹) ^x
'Emphasis'	1 (water)	0.30 b ^w	0.022 b	37.39	30.16	0.059
	1	0.28 b	0.023 b	35.42	28.72	0.067
	7	0.35 b	0.060 a	37.19	21.41	0.065
	14	0.47 a	0.061 a	42.49	22.92	0.062
	21	0.53 a	0.080 a	44.55	24.75	0.070
<i>P</i> value		**	*	NS	NS	NS
'Carnivor'	1 (water)	0.51 b	0.036 d	46.42	27.69 b	0.064
	1	0.49 b	0.042 d	42.53	26.67 b	0.065
	7	0.60 b	0.075 c	46.35	32.17 a	0.059
	14	0.67 ab	0.099 b	48.54	32.66 a	0.060
	21	0.81 a	0.127 a	51.77	33.77 a	0.069
<i>P</i> value		*	***	NS	**	NS

^zDays after treatment [rootstocks were individually treated with 20 μ L (0.0007 fl oz) of either a 6.25% emulsion of fatty alcohol or water].

^ySingle cotyledon measurements.

^x1 g = 0.0353 oz, 1 mm = 0.0394 inch, 1 cm²·g⁻¹ = 4.3942 inch²/oz.

^wMeans within cultivar and column having the same or no letters are not significantly different by Fishers' protected least significant difference test; NS, *, **, *** (not significant or significant at $P \leq 0.05$, 0.01, and 0.001).

Table 3. Effect of time after water and fatty alcohol treatment on ‘Emphasis’ bottle gourd and ‘Carnivor’ interspecific hybrid squash rootstock total soluble sugars (TSS) and starch content per hypocotyl and cotyledon.

Rootstock	DAT ^z	Hypocotyl ^y		Cotyledon ^v	
		TSS ($\mu\text{g}\cdot\text{g}^{-1}$) ^x	Starch ($\mu\text{g}\cdot\text{g}^{-1}$) ^w	TSS ($\mu\text{g}\cdot\text{g}^{-1}$)	Starch ($\mu\text{g}\cdot\text{g}^{-1}$)
‘Emphasis’	1 (water)	1.017 bc ^u	0.023	0.663 b	0.032 c
	1	0.841 c	0.027	0.553 b	0.007 c
	7	1.021 bc	0.185	3.249 ab	1.601 b
	14	1.932 ab	0.469	4.530 a	2.266 b
	21	2.521 a	0.724	5.930 a	3.487 a
P value		*	NS	*	**
‘Carnivor’	1 (water)	1.625 c	0.040 d	1.323 c	0.061 c
	1	1.546 c	0.047 d	1.521 c	0.076 c
	7	4.035 bc	2.440 c	5.322 b	2.218 b
	14	7.763 ab	4.906 b	6.481 ab	2.969 ab
	21	10.824 a	7.742 a	7.620 a	4.140 a
P value		**	**	**	**

^zDays after treatment [rootstocks were individually treated with 20 μL (0.0007 fl oz) of either a 6.25% emulsion of fatty alcohol or water].

^yMean of 12 samples, each sample consisting of five individual hypocotyls.

^x1 $\mu\text{g}\cdot\text{g}^{-1}$ = 1 ppm.

^wMicrograms starch per gram tissue dry weight.

^vMean of 12 samples, each sample consisting of 10 individual cotyledons.

^uMeans within cultivar and column having the same or no letters are not significantly different by Fishers’ protected least significant difference test; ns, *, ** (not significant or significant at $P \leq 0.05$ and 0.01).

starch content of rootstock hypocotyls and cotyledons 1 DAT. Thus, fatty alcohol application itself does not affect baseline rootstock development or carbohydrate storage.

Hypocotyl length of ‘Carnivor’ decreased as time after treatment progressed, which seems improbable, as hypocotyls do not decrease in length once a set length is reached. Further investigation is necessary to verify this result. With the exception of ‘Carnivor’ cotyledons, the increases in hypocotyl and cotyledon size observed over the 21-d experiment agree with the findings of Havis (1940), who reported an increase in cotyledon size and thickness of brassica seedlings after removal of the epicotyl. Similarly, Saks and Ilan (1984) also reported an increase in sunflower (*Helianthus annuus*) cotyledon fresh weight over 18 d after decapitation.

Many studies have reported on the carbohydrate changes in physiologically decapitated woody plants, each with varying results regarding carbohydrate content responses to meristem removal. Tschaplinski and Blake (1994) reported significant decreases after 10 d in both starches and soluble sugars of decapitated hybrid poplar (*Populus maximowiczii* \times *P. nigra*). Another study of defoliated cranberry (*Vaccinium*

macrocarpon) uprights showed no effect of defoliation on total non-structural carbohydrates after 7 d (Vanden Heuvel, 2004). In decapitated australian oak (*Eucalyptus obliqua*) seedlings, soluble sugars decreased by half after 10 d, and then increased to normal levels 40 d after decapitation (Taylor et al., 1982).

Tschaplinski and Blake (1994) attribute the loss of total nonstructural carbohydrates to the loss of leaves, the source of carbon and growth hormones in the plants’ source–sink relationship. In a similar way, watermelon rootstock seedlings may be dependent on the cotyledons. Bisognin et al. (2005) demonstrated the reliance of cucumber (*Cucumis sativus*) seedlings on both cotyledons in carbohydrate accumulation. Rootstock cotyledons are photosynthetic, thereby providing a carbon source for the seedling, which may be adequate for early growth. The importance of the cotyledons, and their retention in our experiment compared with the previously discussed decapitation experiments, may explain the increases in carbohydrate content observed in this study.

The increase in hypocotyl carbohydrate content after fatty alcohol treatment could benefit the watermelon grafting industry. First, rootstock

treated with fatty alcohol may have an increased grafting window. That is, cucurbit rootstock seedlings typically have a narrow window of 1 to 2 d in which they may successfully be grafted without a developing meristem. The fatty alcohol treatment described herein could increase the grafting window up to three weeks, allowing treated plants to be left in the greenhouse to accumulate carbohydrates until ready to be grafted. Especially as increased hypocotyl width may play a key role in graft survival, further studies are required to determine whether the carbohydrate content will affect grafting success. Furthermore, the increase in hypocotyl carbohydrate content could also allow for the elimination of the rootstock cotyledon in grafting, which would decrease grafting costs by allowing for the use of a smaller tray size and smaller grafting space. This type of grafting method, typically used in tomato (*Solanum lycopersicum*) grafting, is used to eliminate the problem of rootstock regrowth, but has not been feasible in cucurbit grafting because of the strong dependence of the rootstock upon the cotyledons. The increase in carbohydrates stored in the hypocotyl may provide sufficient energy to support a cotyledon-devoid graft in cucurbits.

Based on the findings in this study, we conclude that cotyledons and hypocotyls of rootstock seedlings continue to develop and expand over 21 d after fatty alcohol treatment. Cotyledons and hypocotyls also increase in carbohydrate content, most notably starch. This increase in stored energy could allow for increased efficiency of current production practices, as well as new grafting methods that should reduce the cost of grafted transplant production.

Literature cited

Bisognin, D.A., L. Velasquez, and L. Widders. 2005. Cucumber seedling dependence on cotyledonary leaves for early growth. *Pesquisa Agropecu. Bras.* 40:531–539.

Buller, S., D. Inglis, and C. Miles. 2013. Plant growth, fruit yield and quality, and tolerance to verticillium wilt of grafted watermelon and tomato in field production in the Pacific Northwest. *HortScience* 48:1003–1009.

- Choi, D.C., S.W. Kwon, B.R. Ko, and J.S. Choi. 2002. Using chemical controls to inhibited axillary buds of *Lagermaria* as rootstock for grafted watermelon (*Citrullus lanatus*). *Acta Hort.* 588:43–48.
- Cohen, R., Y. Burger, C. Horev, A. Koren, and M. Edelstein. 2007. Introducing grafted cucurbits to modern agriculture: The Israeli experience. *Plant Dis.* 91:916–923.
- Daley, S. and R.L. Hassell. 2014. Fatty alcohol application to control meristematic regrowth in bottle gourd and interspecific hybrid squash rootstocks used for grafting watermelon. *HortScience* 49:260–264.
- Davis, A.R., P. Perkins-Veazie, Y. Sakata, S. Lopez-Galarza, J.V. Marot, S. Lee, Y. Huh, Z. Sun, A. Miguel, S.R. King, R. Cohen, and J. Lee. 2008. Cucurbit grafting. *Crit. Rev. Plant Sci.* 27:50–74.
- Guan, W., X. Zhao, R. Hassell, and J. Thies. 2012. Defense mechanisms involved in disease resistance of grafted vegetables. *HortScience* 47:164–170.
- Hassell, R.L., F. Memmott, and D.G. Liere. 2008. Grafting methods for watermelon production. *HortScience* 43:1677–1679.
- Havis, A.L. 1940. Developmental studies with *Brassica* seedlings. *Amer. J. Bot.* 27:239–245.
- Lee, J. and M. Oda. 2003. Grafting of herbaceous vegetable and ornamental crops. *Hort. Rev.* 28:61–124.
- Louws, F.J., C.L. Rivard, and C. Kubota. 2010. Grafting fruiting vegetables to manage soilborne pathogens, foliar pathogens, arthropods and weeds. *Sci. Hort.* 127:127–146.
- Memmott, F.D. and R.L. Hassell. 2010. Watermelon (*Citrullus lanatus*) grafting method to reduce labor cost by eliminating rootstock side shoots. *Acta Hort.* 871:389–394.
- Penny, M.G., K.G. Moore, and P.H. Lovell. 1976. The effect of inhibition of cotyledon photosynthesis on seedling development in *Cucumis sativus* L. *Ann. Bot. (Lond.)* 40:815–824.
- Rutledge, A.D. 2009. Growing vegetable transplants in Tennessee. *Univ. Tennessee Agr. Ext. Serv.* PB 819. 20 Feb. 2013. <<https://utextension.tennessee.edu/publications/Documents/PB819.pdf>>.
- Saks, Y. and I. Ilan. 1984. Hormone-mediated regulative action of the sunflower shoot apex on growth and cation level in the cotyledons. *Plant Physiol.* 74:408–412.
- Steffens, G.L., T.C. Tso, and D.W. Spaulding. 1967. Fatty alcohol inhibition of tobacco axillary and terminal bud growth. *J. Agr. Food Chem.* 15:972–975.
- Taylor, J.S., T.J. Blake, and R.P. Pharis. 1982. The role of plant hormones and carbohydrates in the growth and survival of coppiced *Eucalyptus* seedlings. *Physiol. Plant.* 55:421–430.
- Tschaplinski, T.J. and T.J. Blake. 1994. Carbohydrate mobilization following shoot defoliation and decapitation in hybrid poplar. *Tree Physiol.* 14:141–151.
- U.S. Department of Agriculture. 2012. Crop values 2011 summary. 15 Apr. 2012. <<http://usda01.library.cornell.edu/usda/current/CropValuSu/CropValuSu-02-16-2012.pdf>>.
- Vanden Heuvel, J.E. 2004. Source-sink relationships in cranberry: Effects on carbohydrate production and partitioning. *HortScience* 39:761 (abstr.).
- Zhao, D., C.T. MacKown, P.J. Starks, and B.K. Kindiger. 2010. Rapid analysis of nonstructural carbohydrate components in grass forage using microplate enzymatic assays. *Crop Sci.* 50:1537–1545.