

Figure 1. Processes potentially affected by antibiotics in soil.

INTRODUCTION

Animal excrement and wastewater often contain trace concentrations of un-metabolized veterinary and human antibiotic compounds. When these are used in agriculture for fertilizer and/or irrigation, these compounds have the potential to affect microbial activity in soils. The objective of this research is to evaluate the impact of trace Narasin exposure on nitrification, denitrification, and N₂O flux rates.



Figure 2. Soil incubation chambers equipped with headspace sampling ports.

EXPERIMENTAL METHODS

In replicates of six, 75 g air-dried soil (sandy loam) was treated with 0-1000 ng/kg Narasin and a 50:50 mixture of KNO₃:(NH₄)₂SO₄ in amounts equivalent to 100 kg N hectare⁻¹. At each dose, 3 samples were enriched with ¹⁵N-NO₃⁻ and 3 with ¹⁵N-NH₄⁺ (10% atom excess). Total volumetric water content was brought to 40% Water-Filled Pore Space (Experiment 1) and 60% Water Filled Pore Space (Experiment 2) and the soils were placed in gas-tight jars (Figure 2) and allowed to incubate for 3 days at room temperature. Soil extracts (2M KCl) and headspace samples were collected daily and used to quantify NO₃, NH₄⁺, and N₂O flux.

EFFECTS ON MINERAL NITROGEN

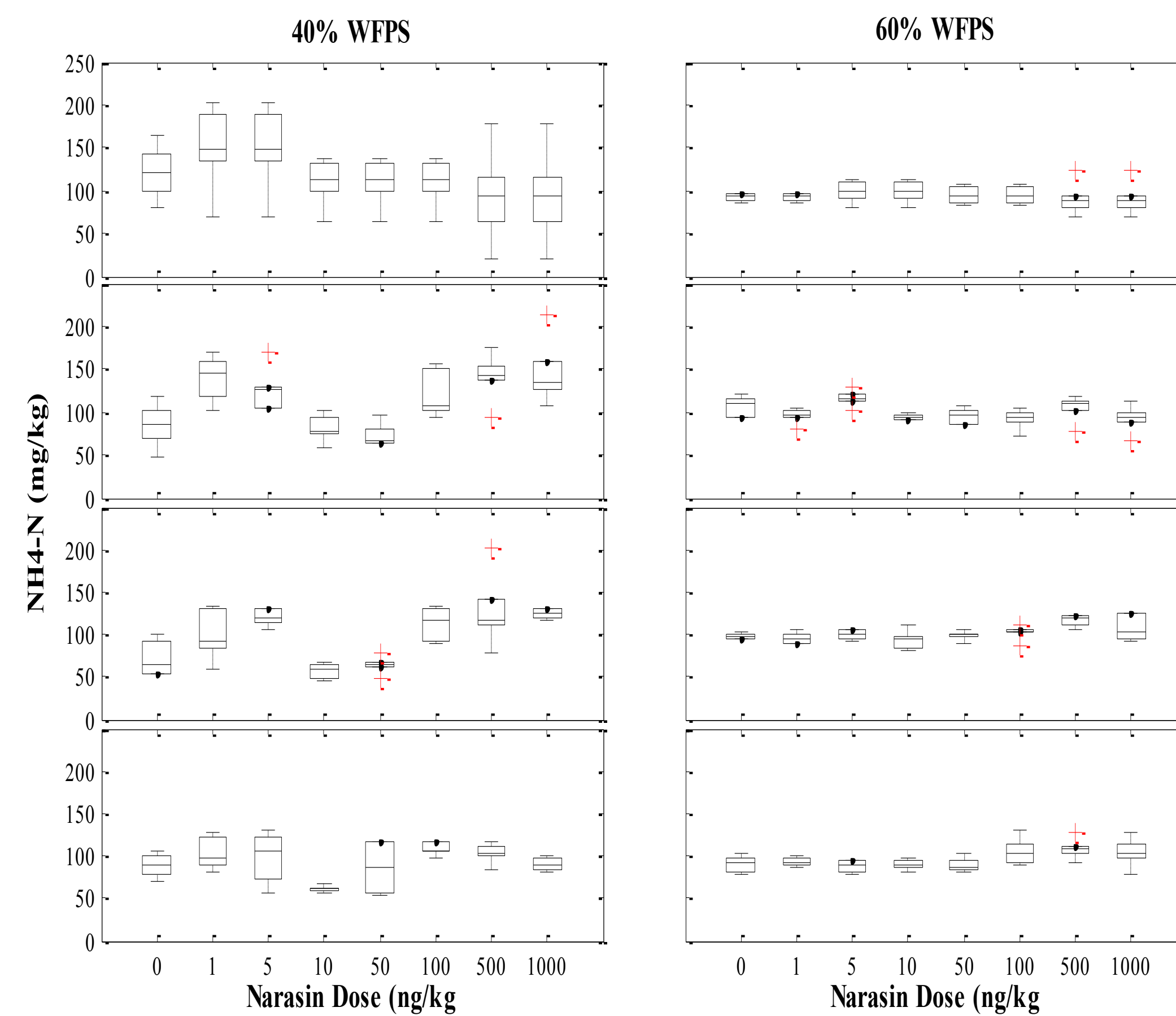


Figure 3. Box-whisker plots illustrating extractable NH₄⁺ over the course of soil incubations. At 40% WFPS, a statistically significant dose response was observed on Day 1 ($p = 2.58E-06$), Day 2 ($p = 1.3E-07$), and Day 3 ($p = 1.66E-03$) where the concentration of NH₄⁺ in treated soils exceeded that of untreated soils at all but the 10 and 50 ng kg⁻¹ Narasin doses. At 60% WFPS, a statistically significant dose response on Day 1 ($p = 0.00$), Day 2 ($p = 0.0005$), and Day 3 ($p = 0.004$). Here, NH₄⁺ was depleted relative to the control at all doses on Day 1 but equal to or in excess of the control on Days 2 and 3.

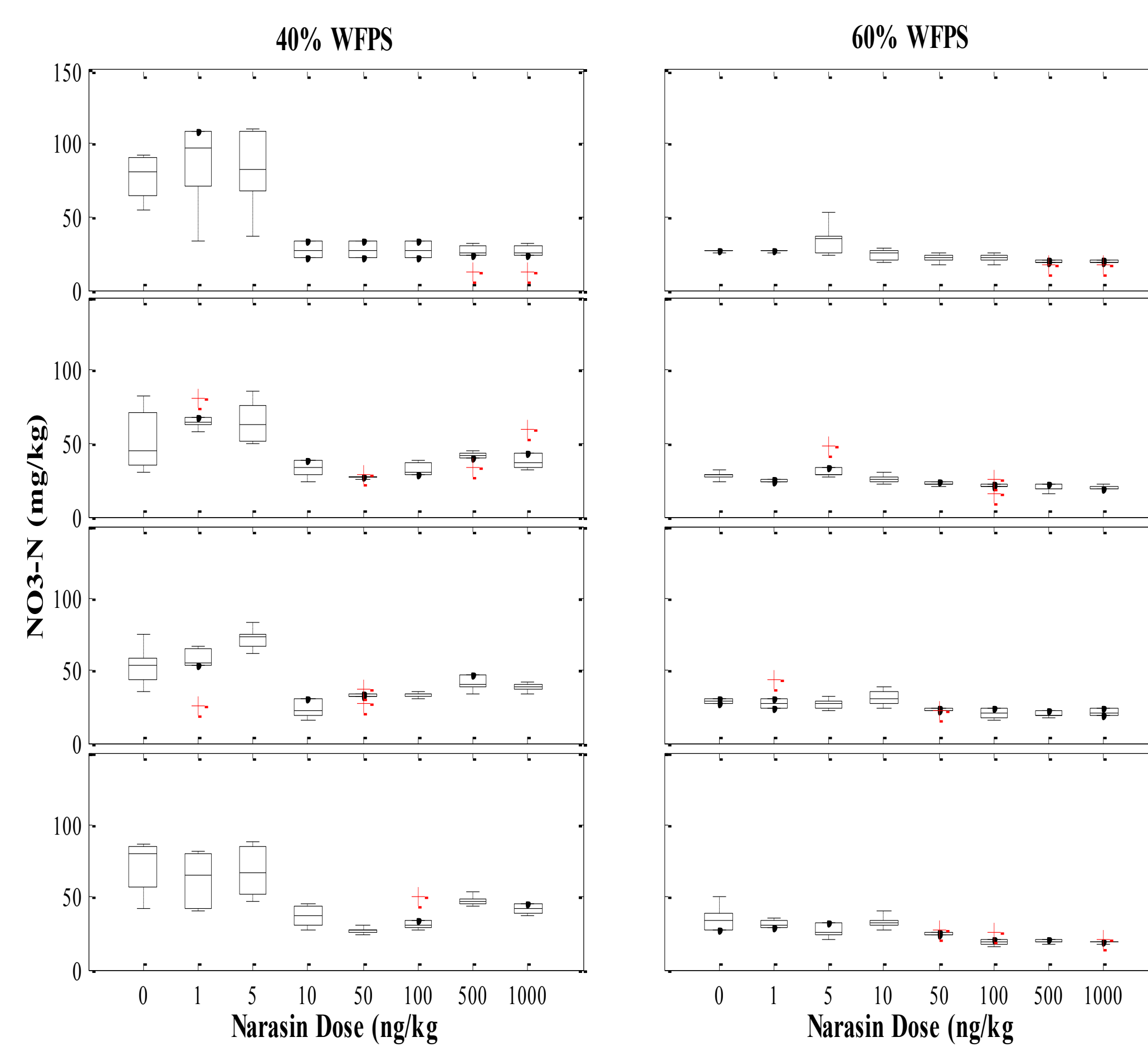


Figure 4. Box-whisker plots illustrating extractable NH₄⁺ over the course of soil incubations. At 40% WFPS, a statistically significant dose response was observed on Day 1 ($p = 4.83E-08$), Day 2 ($p = 1.2E-11$), and Day 3 ($p = 2.34E-08$) where the concentration of NO₃⁻ in treated soils was depleted relative to the control at Narasin doses greater than 10 ng kg⁻¹. A more distinctly linear pattern in which increasing dose correlates to decreased NO₃⁻ was observed at 60% WFPS, with statistically significant dose-responses observed on Day 1 ($p = 6.62E-07$), Day 2 ($p = 2.34E-06$), and Day 3 ($p = 3.62E-09$).

EFFECTS ON N₂O FLUX

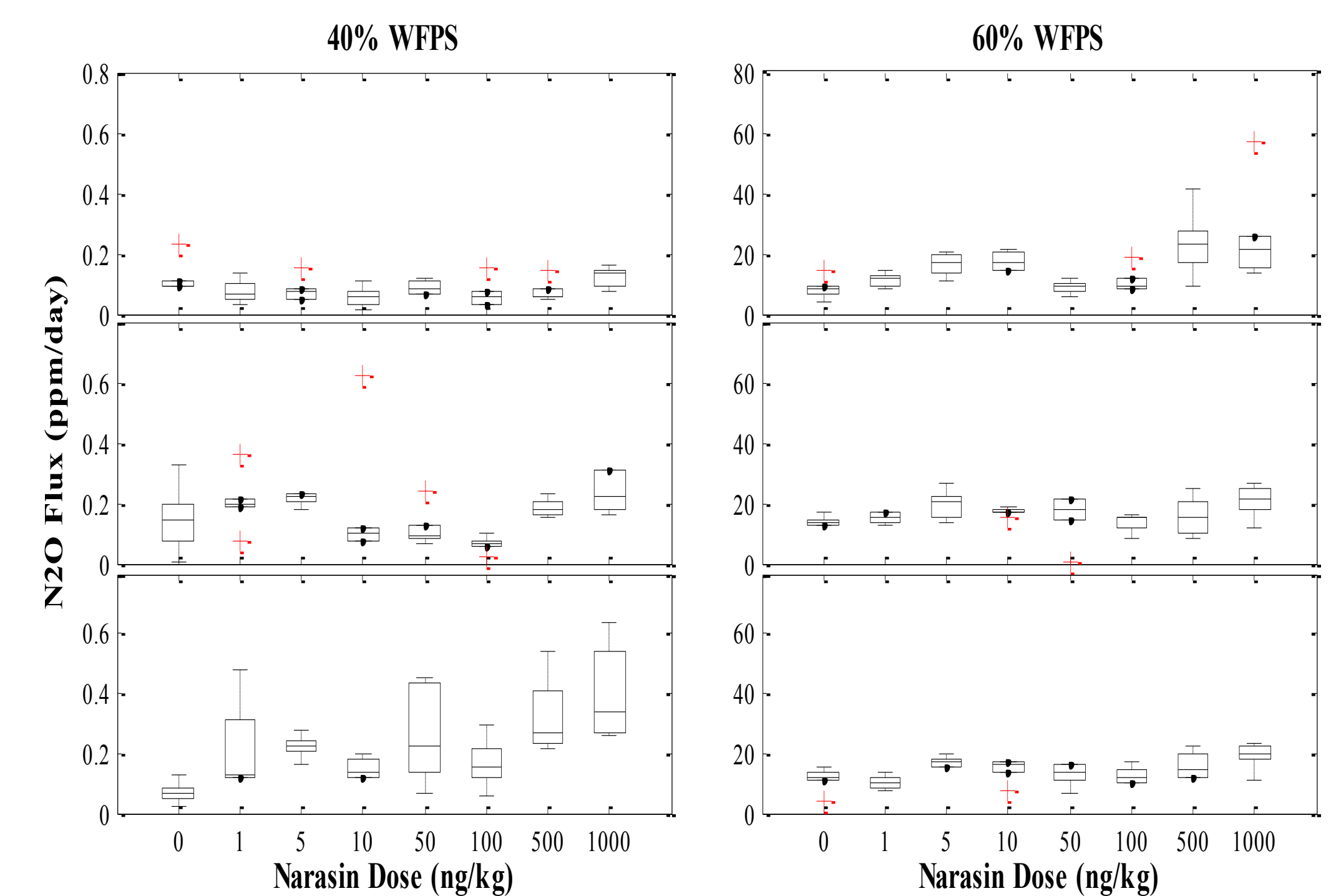


Figure 5. Box-whisker plots illustrating N₂O flux (ppm/day). At both 40% and 60% WFPS, increasing antibiotic doses correlate to increasing N₂O flux rates. At the lower water content, the significance of this effect is delayed until Day 3 ($p = 0.006$) whereas the pattern is immediately observed in soils raised to 60% WFPS ($p = 0.0006$), muted on Day 2 ($p = 0.115$), and observed again on Day 3 ($p = 0.0007$).

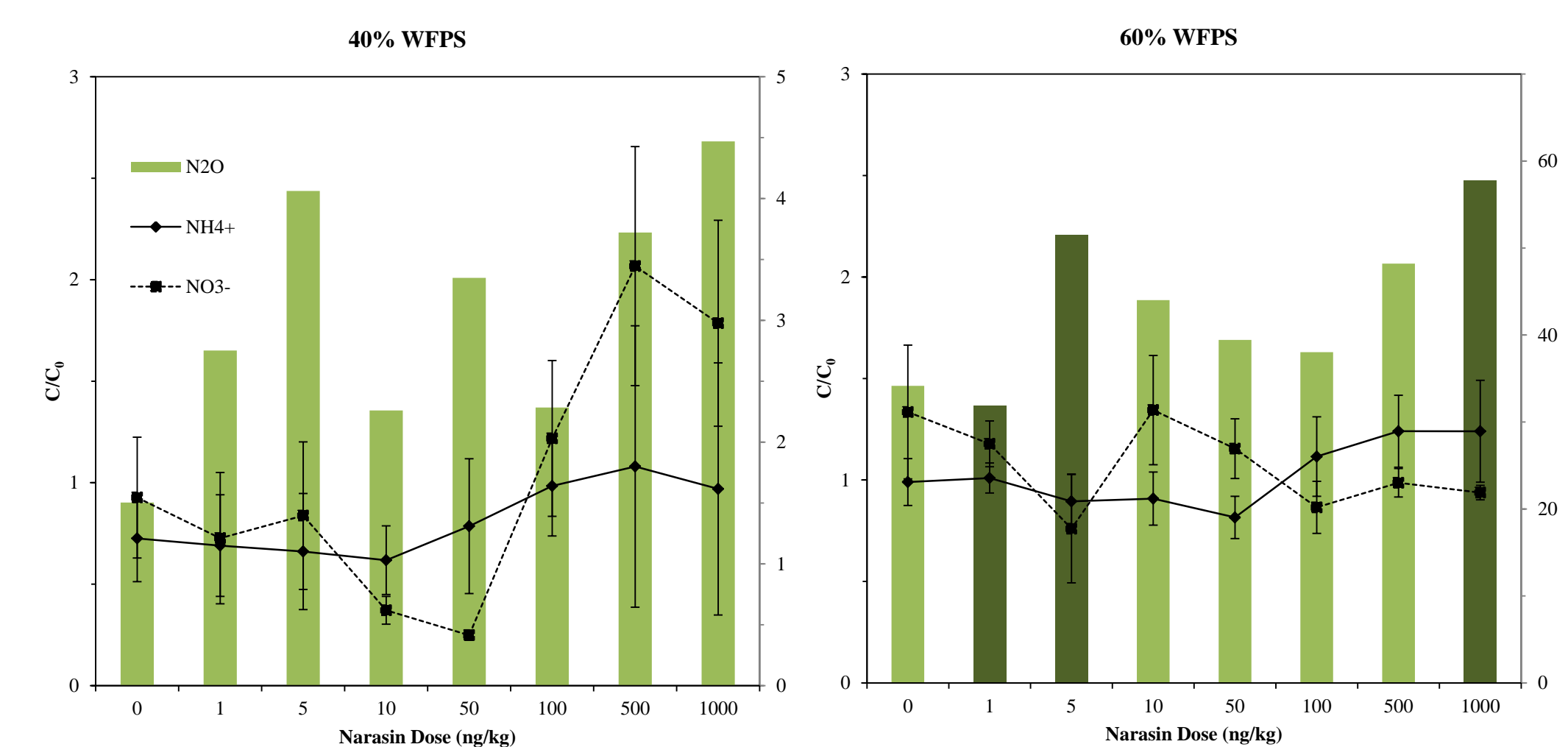


Figure 6. A comparison of mineral nitrogen on Day 3 (C/C₀) to total N₂O flux shows little appreciable correlation between mineral N species and N₂O flux rate in drier soils, but NO₃⁻ and N₂O show similar trends at 60% WFPS for all but the 500 and 1000 ng kg⁻¹ doses.

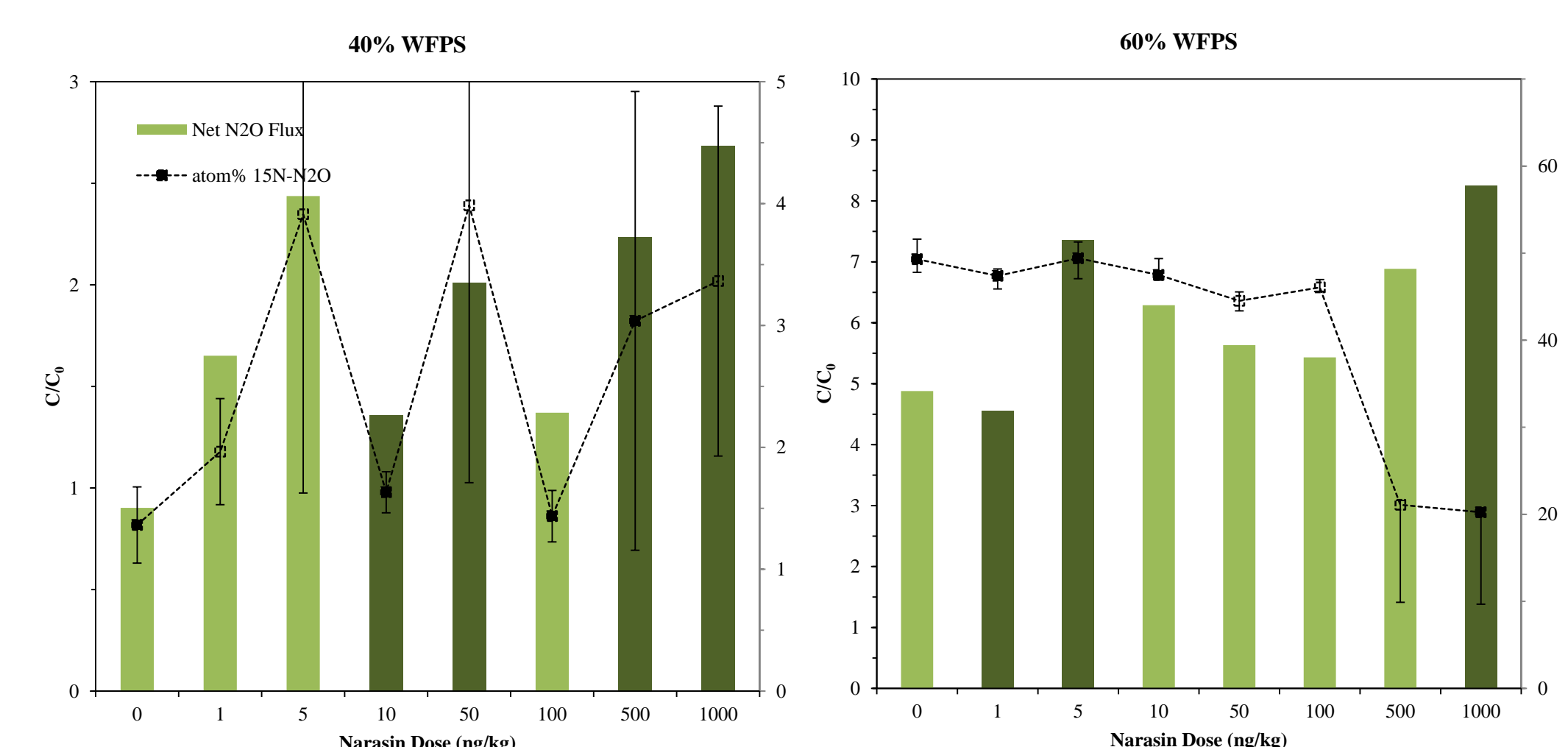


Figure 7. The ¹⁵N isotopic enrichment of N₂O from soils that were treated with ¹⁵N-KNO₃ strongly correlate to total N₂O accumulation on Day 3, suggesting that denitrification is the primary N₂O source. The distinct disagreement between ¹⁵N-N₂O and total N₂O flux at 60% WFPS, 500 and 1000 ng kg⁻¹ doses implies significant dilution of the NO₃⁻ pool by mineralization and nitrification, coupled with denitrification.