Characterization of Agouti-Signaling Protein and its Receptors in the Bovine Oocyte and Early Embryo

Heather L. Chaney, Jaelyn Z. Current, Mingxiang Zhang, and Jianbo Yao

Davis College of Agriculture, Natural Resources and Design, Division of Animal and Nutritional Sciences, West Virginia University, Morgantown, WV, USA

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Introduction

Bovine *In Vitro* Fertilization (IVF)

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- For the first time, it was reported a greater number of bovine embryos were generated via IVF than in vivo in 2016 [1].
- Despite advancements in oocyte and embryo culture systems, only 20-40% of presumptive zygotes will reach the blastocyst stage in vitro [2].
- A limiting factor to IVF is lack of knowledge of the mRNAs and proteins necessary for oocyte maturation and early embryonic development [3,4].

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Agouti signaling protein (ASIP):	700 -	
 132 amino acid secreted protein 	600 -	
• Highly abundant in the bovine oocyte (RNA-	VX 500 -	
Sequencing data, Yao lab; Fig. 1).	- 400 - 300 -	

Results



- The function of ASIP in the ovary is unknown.
- in lipid metabolism has been • A role established in other bovine tissues including adipose [5] and mammary epithelial cells [6].
- 200 100 ASIP KPNAT NPM2 NLRP9 2AR1 ACTE
- ASIP binds to melanocortin receptors (MC1R-5R) and the attractin (ATRN) receptor [6].
- Figure 1. ASIP expression in the bovine oocyte in relation to other highly expressed oocyte genes.

Objectives:

- 1. Characterize ASIP expression throughout early embryonic development and within the ovarian follicle.
- 2. Determine the expression of putative receptors for ASIP in follicular cells and the oocyte.

Materials and Methods



Figure 3. Expression of ASIP during oocyte maturation and early embryogenesis. Data are relative to *GFP* RNA levels. Expression was detected in all stages (n = 4 pools of 20 oocytes/embryos) with the highest levels present at the GV and blastocyst stages (P < 0.0001).



Figure 2. Workflow for experiments 1 and 2. Oocytes were obtained from slaughterhouse bovine ovaries and aspirated to obtain cumulus-oocyte complexes (COCs) or antral follicles 3-10 mm were dissected for cell isolation.

In vitro embryo production

- Cumulus-oocyte complexes (COCs) were cultured in IVM (IVF) Bioscience) for 22-24 h and fertilized.
- Presumptive zygotes were placed in IVC (IVF Bioscience), and 2-, 4-, 8-, 16, morula, and blastocyst stage embryos were collected then stored at -80°C until RNA isolation.

Analysis of gene expression via RT-qPCR (Fig. 2)

- RNA was isolated from all samples using the RNAqueous Total RNA Isolation Kit (Invitrogen).
- RNA was reverse transcribed using the High-capacity cDNA Reverse Transcription Kit (Applied Biosystems).
- RT-qPCR was conducted using PowerUp SYBR Green Master Mix (Applied Biosystems) according to published methods [7].
- Gene normalization:

Figure 4. Putative ASIP receptors detected in oocytes (A) and follicular cells (B). (A) Expression of ATRN, MC3R, and MC4R was present in both GV and MII oocytes (n = 4 pools of 10). No statistical difference between stages (GV vs. MII; P > 0.05). (B) ATRN, MC3R, and MC4R were also detected in ovarian follicular cells including cumulus, granulosa, and theca cells isolated form antral follicles (n = 12-16). Cumulus expression of receptors ATRN, MC3R, and MC4R was the highest (P < 0.0001).

Conclusions

- ASIP is expressed in follicular cells including cumulus, granulosa, and theca cells.
- ASIP expression decreases following oocyte maturation and rises following activation of the embryonic genome at the 8-16 cell stage.
- ASIP putative receptors, ATRN, MC3R, and MC4R are expressed in both GV and MII oocytes. ATRN was detected at high levels in cumulus, granulosa, and theca cells while MC3R and MC4R were detected in cumulus and granulosa cells.
- Future work will characterize the function of ASIP during oocyte maturation and early embryonic development in cattle.
- Embryo panel samples were spiked with GFP RNA during RNA isolation, which was used for normalization.
- *RPL19* expression was utilized for normalization of all other data.
- Relative expression values were calculated using the standard curve method.
- Data are presented as mean \pm SEM.
- Log-transformed RT-qPCR data were used for statistical analysis.
- Statistical analysis included either a Student's t-test (Fig. 4A) or one-way ANOVA followed by Tukey's HSD.
- $P \le 0.05$ considered significant.

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References

Ferré LB, Kjelland ME, Strøbech LB, Hyttel P, Mermillod P, Ross PJ. Review: Recent advances in bovine. Animal 2020; 14:991-1004. Lonergan P, Fair T, Forde N, Rizos D. Embryo development in dairy cattle. Theriogenology 2016; 86:270-277 3. Sirard MA, Richard F, Blondin P, Robert C. Contribution of the oocyte to embryo quality. Theriogenology 2006; 65:126-136 4. Patel OV, Bettegowda A, Ireland JJ, Coussens PM, Lonergan P, Smith GW. Functional genomics studies of oocyte competence: evidence that reduced transcript abundance for follistatin is associated with poor developmental competence of bovine oocytes. Reproduction 2007; 133:95-106. 5. Liu Y, Albrecht E, Schering L, Kuehn C, Yang R, Zhao Z, Maak S. Agouti Signaling Protein and Its Receptors as Potential Molecular Markers for Intramuscular and Body Fat Deposition in Cattle. Frontiers in Physiology 2018; 9. 5. Xie T, Liu Y, Lu H, Iqbal A, Ruan M, Jiang P, Yu H, Meng J, Zhao Z. The Knockout of the ASIP Gene Altered the Lipid Composition in Bovine Mammary Epithelial Cells via the Expression of Genes in the Lipid Metabolism Pathway. Animals (Basel) 2022; 12. Hand JM, Zhang K, Wang L, Koganti PP, Mastrantoni K, Rajput SK, Ashry M, Smith GW, Yao J. Discovery of a novel oocyte-specific Kruppel-associated box domain-containing zinc finger protein required for early embryogenesis in cattle. Mech Dev 2017; 144:103-112.



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